



pISSN 1225-8318 eISSN 2466-1546

RESEARCH ARTICLE https://doi.org/10.11110/kjpt.2023.53.4.271

New diploid populations of the Chrysanthemum zawadskii complex (Asteraceae) in natural habitats of Korea

Mi Jeong MOON, Sang Hee PARK, Sung Kyung HAN¹ and Jung Sung KIM*

Department of Forest Science, Chungbuk National University, Cheongju 28644, Korea ¹Baekdudaegan Bioresources Conservation Department, Baekdudaegan National Arboretum, Bonghwa 36209, Korea

Corresponding author Jung Sung KIM E-mail: jungsung@cbnu.ac.kr



© 2023 The Author(s) This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (Ihtp://creativecommons.org/licenses/by-nc/ 4.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

ABSTRACT: Chrysanthemum zawadskii Herbich is widely distributed from northern Europe to East Asia. It has been reported that various ploidy levels ranging from diploid (2n = 2x =18) to decaploid (2n = 10x = 90) are distributed in Korea. The diploid populations of C. zawadskii, however, have limited habitats, with only four populations from two locations compared to polyploids that are widely distributed in Korea. During recent biodiversity research on the C. zawadskii complex, new habitats of diploid populations of C. zawadskii not reported earlier in Korea were identified. To understand the speciation mechanism accompanying polyploidization of the C. zawadskii complex, it was necessary to investigate the features of these new populations as to whether they identical to or different from previously reported diploid populations. For this purpose, we identified the number and karyotypes of somatic chromosomes in four new diploid populations and compared the morphological features. It was found that the new diploid populations differed slightly in terms of the size and color of the flowers and in the shape of the leaves, with continuous variations. Representative karyotypes of the new populations were included in previously reported diploid karyotypes, but the numbers and locations of satellite chromosomes differed. In addition, we found that one of them is a sympatric population with different ploidy levels, including diploid, tetraploid, and hexaploid. From these results, it was confirmed that morphological and cytological variations exist among diploid populations in the C. zawadskii complex.

KEYWORDS: chromosome number, *Chrysanthemum zawadskii* complex, diploid, karyo-typing, ploidy

RECEIVED 3 August 2023; REVISED 9 November 2023; ACCEPTED 22 December 2023

INTRODUCTION

Within the Asteraceae family, genus *Chrysanthemum* L., a well-known genus possessing various cultivars, is distributed primarily in Eastern Asia and the temperate climate zone of Siberia (Kadereit and Jeffrey, 2007; Plants of the World Online, 2019). About the circumscription of the species of the genus *Chrysanthemum*, many researchers have traditionally argued and proposed several classification systems to identify regional wild *Chrysanthemum* (Makino, 1903; Hu, 1958; Kitamura, 1978; Anderson, 1987; Trehane, 1995; Kim et al., 2003). Korean *Chrysanthemum*, is generally classified into three species of *Chrysanthemum zawadskii* complex, *Chrysanthemum indicum* L., and *Chrysanthemum boreale* Makino. The former

C. zawadskii have ray flowers of white or pale pink, unlike the latter two species which have yellow ray flowers (Lee, 2003, 2006; Kim et al., 2014). However, the genus *Chrysanthemum* has a wide range of morphological variations, particularly in the size, shape, and anatomical structure of leaves, and these variations result in the ambiguity of the species circumscription of the genus (Oh et al., 1994; Kim and Tobe, 2009; Kim et al., 2014; Kang and Kim, 2020).

The basic chromosome number of the genus *Chrysanthemum* is x = 9, which indicates a wide range of ploidy (Liu et al., 2012). To date, a series of polyploids: diploids (2n = 2x = 18), tetraploids (2n = 4x = 36), pentaploids (2n = 5x = 45), hexaploids (2n = 6x = 54), and decaploids (2n = 10x = 90), have been reported in the Korean *Chrysanthemum* (Lee, 1967,

1969; Kim et al., 2003, 2008). In case of *Chrysanthemum coreanum* (H. Lév. & Vaniot) Nakai, a member of the *C. zawadskii* complex restricted to Mt. Hallasan on Jejudo Island, has been confirmed to be a decaploid (10x = 90) and not an octoploid (2n = 8x = 72) (Kim et al., 2004). Because ploidy has regarded as a vital factor related to plant evolution (Leitch and Bennett, 1997; Te Beest et al., 2012), the discovery of various ploidy populations has important implications for the study of the genus *Chrysanthemum*. Especially, the presence or absence of diploid populations can play a role in resolving plant evolution via polyploidy. The precise confirmation of ploidy levels at the population scale is crucial to accurately understanding the speciation and evolutionary history of the genus *Chrysanthemum*.

For a very long time, chromosome numbers have been used as a cytogenetic characteristic to explain the most basic genetic properties of living organisms (Mayrose and Lysak, 2021), and karyotypes include the number, shape, and size of chromosomes, have been generally used as one of important tools for species identification (Heslop-Harrison and Schwarzacher, 2011). The chromosome-based researches have been conducted on various species of the genus *Chrysanthemum*, especially the Japanese, Chinese, and Taiwanese species (Tanaka and Shimotomai, 1961; Wang et al., 1993; Kondo et al., 1999; Kim et al., 2004), and has provided us with cytogenetic insights to understand the complex relationships among the species and different ploids.

Chrysanthemum zawadskii Herbich is the only widespread species in the genus reported from Europe to Asia, and it forms a species complex of different ploidy levels and various infra-specific taxa in Korea although its correct taxonomical status is still being debated. Considering the first discoveries in the diploid populations of this species (sites A and B) (Fig. 1) (Kim et al., 2003), the successive ploids from diploid to decaploid were completed, and increasing the probability of understanding the evolutionary process in the genus. However, the number of diploid populations is still restricted than that of the tetra- and hexaploid populations which are widely distributed across Korea. Therefore, it still makes a limitation for the discussion on the diversity of the *C. zawadskii* complex as well as the genus *Chrysanthemum*.

During our recent investigation, we identified new populations of diploid *C. zawadskii* native to Korea. In this study, we describe the morphological and cytological characteristics of these new populations along with their relationships.



Fig. 1. Distribution map of the newly discovered diploid populations of *Chrysanthemum zawadskii* in Korea. The details of each population are presented in Table 1. **A** and **B** on the map indicate previously reported diploid population sites by Kim et al. (2003). **A.** Gwangha-ri of Jeongseon-gun and Baekun-ri of Pyeongchang-gun, Gangwon-do. **B.** Dalsan and Okgye of Yeongdeok-gun, Gyeongsangbuk-do.



Fig. 2. The native habitats of the new diploid populations of *Chrysanthemum zawadskii*. A. Mt. Seokbyeongsan. B. Mt. Gakhuisan. C. Mt. Baekisan. D. Miin Falls.

Table 1. Information on the collection of the new diploid Chrysanthemum zawadskii populations in Korea.

Population	Living material No.	Locality	Coordination	Elevation (m)	Chromosome number $(2n)$
Mt. Seokbyeongsar	CBNU2020LM-0631	Gangwon-do, Gangneung-si, Okgye-myeon, Sangye-ri, Mt. Seokbyeongsan	37°35'12"N, 128°53'51"E	994	18
Mt. Gakhuisan	CBNU2022LM-0327	Gangwon-do, Jeongseon-gun, Hwaam-myeon, Hwaam-ri, Mt. Gakheesan	37°21'22"N, 128°48'20"Е	930	18
Mt. Baekisan	CBNU2023LM-0001	Gangwon-do, Jeongseon-gun, Nam-myeon, Gwangdeok-ri, Mt. Baekisan	37°17'45"N, 128°42'12"E	563	18
Miin Falls	CBNU2022LM-0335	Gangwon-do, Samcheok-si, Dogye-eup, Simpo-ri, Miin Falls	37°10'42"N, 129°02'48"E	700	18

MATERIALS AND METHODS

Plant materials and morphology observation

Four wild populations of *C. zawadskii* were identified by morphological characteristics as described by Lee (2003, 2006) and were collected from the top or ridge of a mountain before flowering season (except for one population; Miin Falls was collected after flowering) in 2022 in Korea (Figs. 1, 2, Table 1). All the collected materials were transplanted and nurtured as living samples in a greenhouse at Chungbuk National University (Cheongju-si, Chungbuk, Korea) to confirm their morphological and chromosomal characteristics. A total of eight characteristics, including head flower size, color, and leaf size were investigated and measured from more than three individuals per population based on their flowering conditions to compare the morphology among the populations (Table 2).

Fixation of the root tip

According to Kim et al. (2003), healthy root tips were cut approximately 2–3 cm from the living material. The root tips were soaked in 10 mL 0.002 M 8-hydroxyquinoline solution for 2 h at room temperature (RT) for the pretreatment, and then it was carried out in a fixative solution (99% acetic acid:99.9% ethanol = 1:3 v/v) for 30 min on ice. The fixed roots were stored in 70% ethanol (99.9% ethanol:distilled water = 7:3 v/v) at -20° C until chromosome observation.

Chromosome observations and karyotype analysis

According to the protocols of Fan (1965) and Kim et al. (2003), the fixed root tip was dissociated for 90 s at 65°C in 1N HCl and placed on a slide glass. After being crushed, the root tip tissue was then separated and stained using 1% acetoorcein for 35 min at RT in a humid chamber. The mitotic metaphase chromosomes were observed under an optical microscope (Olympus BX50; Tokyo, Japan). To get the representative karyotype of each population, total of four individuals were randomly selected from new diploid populations and at least three slides per individual were made for observation. Finally, over three cells in the mitotic metaphase stage were analyzed for karyotyping per slide.

For karyotype analyses, high-quality magnified images were captured and edited using Adobe Photoshop CC (Adobe Systems, San Jose, CA, USA). The chromosome length was measured using KaryoMeasure software (Mahmoudi and Mirzaghaderi, 2023), and the karyotype was analyzed as per the standards set suggested by Levan et al. (1964). Based on the ratio of short arm to long arm and the centromere locations of the chromosome, the karyotypes were identified and

Table 2. Morphological characteristics of the new diploid Chrysanthemum zawadskii populations in Korea.

1 0			1 2		1 1			
Population	Flower color (RHS)	Head diameter (mm)	Disk floret width (mm)	Ligulate length (mm)	Leaf length (cm)	Leaf width (cm)	L/W ratio	Petiole length (cm)
Mt. Seokbyeongsan	NN155C (White)	28.9 ± 6.30	9.0 ± 0.86	21.4 ± 6.35	3.0 ± 0.42	3.1 ± 0.33	1.0 ± 0.05	3.7 ± 0.70
Mt. Gakhuisan	85C (Very Light purple)	35.0 ± 1.00	9.0 ± 0.93	26.0 ± 1.93	2.8 ± 0.54	2.7 ± 0.29	1.0 ± 0.09	3.0 ± 0.42
Mt. Baekisan	NN155C (White)	29.9 ± 2.67	9.7 ± 0.93	20.2 ± 2.40	3.6 ± 0.29	3.7 ± 0.29	1.0 ± 0.11	3.6 ± 0.57
Miin Falls	NA	NA	NA	NA	2.0 ± 0.05	2.0 ± 0.00	1.0 ± 0.00	1.5 ± 0.38

L/W, length/width; NA, not available.



Fig. 3. Comparison of morphological characteristics among the new diploid populations of *Chrysanthemum zawadskii*; the upper row: the head flower of Mt. Seokbyeongsan (A), Mt. Gakhuisan (B), and Mt. Baekisan (C). Lower row: the leaves positioned at the upper and middle part of plant collected from Mt. Seokbyeongsan (a), Mt. Gakhuisan (b), Mt. Baekisan (c), and Miin Falls (d). Scale bars = 1 cm.

described as follows: m, metacentric chromosomes; sm, submetacentric chromosomes; st, sub-telocentric chromosomes; t, telocentric chromosomes. In case of the chromosome with satellite, it was added an asterisk in the figure and table.

Flow cytometry analysis

The ploidy levels of the diploid *Chrysanthemum* populations were measured with a tetraploid *Solanum tuberosum* (1C = 1.82 pg) (Valkonen et al., 1994) as a standard reference. In a Petri dish placed in ice, leaves of *Chrysanthemum* individuals and standard reference were chopped with a sharp blade using 500 µL of nuclei extraction buffer (CyStain UV Precise P kit, Sysmex Partec, Munster, Germany). Only the extract was filtered with non-sterile CellTrics Filter Green 30 µm (Sysmex Partec), the nuclei were stained with 2,000 µL staining buffer from the CyStain UV Precise P kit (Sysmex Partec). Ploidy level was measured at least three times per sample and recorded from each sample using CyFlow Cube 6 (Sysmex Partec). In case of the sympatric population Mt. Seokbyengsan, additionally found tetraploid and hexaploid were also measured together with standard reference and diploid populations for genome size estimation.

RESULTS

Morphological characteristics of diploid C. zawadskii

The morphological features of the four newly discovered diploid populations of *C. zawadskii* were compared even though the flower characters of Miin Falls population were not available due to late collection after flowering season. The results confirmed the existence of variations in flower size, color, and leaf shape (Fig. 3, Table 2). The flower heads were light purple-to-white in color. The flowers collected from Mt. Seokbyeongsan were very light purple until the end of the

flowering season, whereas Mt. Gakhuisan and Mt. Baekisan populations had mostly white flowers. The head flower size of plants from the population of Mt. Gakhuisan was bigger than that of the other populations. Although the leaf length and width varied among the four populations, the length/width ratios ranged between 0.9 to 1.1.

Counting chromosome numbers and karyotype analysis

Based on the observations of mitotic metaphase chromosomes, it was confirmed that the four newly found populations were diploid (2n = 2x = 18) (Fig. 4) without exception. The representative karyotype of each population was confirmed (Fig. 5, Table 3) and grouped into three types: (1) 2n



Fig. 4. Mitotic metaphase chromosomes of the new diploid populations (2n = 18) of *Chrysanthemum zawadskii*. **A.** Mt. Seokbyeongsan. **B.** Mt. Gakhuisan. **C.** Mt. Baekisan. **D.** Miin Falls. Scale bar = 10 µm.

Α	K	i۱	15	::	71	"	رد	**	11
в	;+	; }	11	::	11	11	12	11	* *
С	11	11	51	/ 1		13	Ŧ \$	* *	11
D	11	11	11)(()	K	11	ii	* * ((

Fig. 5. Representative karyotypes of the new diploid populations of *Chrysanthemum zawadskii*. **A.** Mt. Seokbyeongsan. **B.** Mt. Gakhuisan. **C.** Mt. Baekisan. **D.** Miin Falls. Scale bar = $10 \mu m$.

T	ab	le	3.	Summary	of	karvotype a	nalysis	for newl	y f	ound	dip	loid	pop	ulations	of	Chi	vsanthemun	ı zawadsk	<i>kii</i> i	n k	Corea
						2 2 1	~		~								~				

	C1	-				
Population	Chr. No.	Long arm (μm)	Short arm (μm)	Total (µm)	Chr. type	Karyotype
Mt. Seokbyeongsan	1	2.80 ± 0.17	2.62 ± 0.05	5.41 ± 0.23	m	$2n = 11m + 5sm + 2st^*$
	2	3.18	1.67 ± 0.02	4.85 ± 0.02	sm	
	3	2.41 ± 0.02	2.38	4.79 ± 0.02	m	
	4	2.41 ± 0.09	2.34 ± 0.03	4.75 ± 0.12	m	
	5	2.59 ± 0.08	1.83 ± 0.07	4.42 ± 0.01	m	
	6	2.37 ± 0.07	2.01 ± 0.03	4.38 ± 0.04	m	
	7	2.36 ± 0.05	1.15 ± 0.14	3.5 ± 0.09	sm	
	8	2.71 ± 0.08	0.74 ± 0.05	3.45 ± 0.03	st*	
	9A	3.02	1.46	4.47	sm	
	9B	2.07	1.67	3.74	m	
Mt. Gakhuisan	1	1.95 ± 0.14	1.76 ± 0.03	3.71 ± 0.11	m	$2n = 12m + 5sm^* + 1st^*$
	2	2.16 ± 0.09	1.13 ± 0.09	3.3 ± 0.18	sm	
	3	1.93 ± 0.10	1.24 ± 0.01	3.16 ± 0.09	m	
	4	1.68 ± 0.01	1.45 ± 0.01	3.1 ± 0.01	m	
	5	1.62 ± 0.03	1.43 ± 0.02	3.04 ± 0.05	m	
	6	1.55 ± 0.01	1.52 ± 0.01	3.06 ± 0.01	m	
	7	1.70 ± 0.08	1.07 ± 0.01	2.77 ± 0.08	m	
	8	1.81 ± 0.07	0.96 ± 0.02	2.78 ± 0.05	sm	
	9A	1.97	0.81	2.78	sm*	
	9B	1.86	0.55	2.41	st*	
Mt. Baekisan	1	2.16 ± 0.10	1.3 ± 0.05	3.76 ± 0.15	m	$2n = 11m + 5sm + 2st^*$
	2	1.95 ± 0.03	1.69 ± 0.01	3.65 ± 0.05	m	
	3	2.25 ± 0.14	1.06 ± 0.05	3.31 ± 0.19	sm	
	4	1.68 ± 0.08	1.63 ± 0.07	3.31 ± 0.15	m	
	5	1.52 ± 0.05	1.46 ± 0.08	2.98 ± 0.13	m	
	6	1.80 ± 0.01	1.16 ± 0.02	2.96 ± 0.03	m	
	7	1.71 ± 0.02	0.64 ± 0.07	2.35 ± 0.07	sm	
	8	1.90 ± 0.00	0.48 ± 0.03	2.38 ± 0.03	st*	
	9A	1.83	0.66	2.49	sm	
	9B	1.93	0.44	2.37	st	
Miin Falls	1	2.31 ± 0.11	1.22 ± 0.05	3.53 ± 0.16	sm	$2n = 10m + 6sm + 2st^*$
	2	1.60 ± 0.01	1.56 ± 0.02	3.16 ± 0.01	m	
	3	1.62 ± 0.01	1.42 ± 0.08	3.04 ± 0.09	m	
	4	1.81 ± 0.01	1.20 ± 0.05	3.01 ± 0.04	sm	
	5	1.95 ± 0.05	1.02 ± 0.04	2.97 ± 0.01	m	
	6	1.96 ± 0.00	1.39 ± 0.01	2.83 ± 0.06	m	
	7	1.96 ± 0.00	1.25 ± 0.08	2.82 ± 0.05	m	
	8	1.93 ± 0.03	0.51 ± 0.03	2.39 ± 0.06	sm	
	9	1.96	0.78 ± 0.01	2.32 ± 0.03	st*	

Asterisk indicates a chromosome with satellite.

= $11m + 5sm + 2st^*$ found on Mt. Seokbyeongsan (pop. 1) and Mt. Baekisan (pop. 3); (2) $2n = 12m + 4sm + 1sm^* + 1st^*$ found on Mt. Gakhuisan (pop. 2), and (3) $2n = 10m + 6sm + 2st^*$ found on Miin Falls (pop. 4). Although the representative karyotypes of pop. 1 and 3 were same, the chromosome shape of 9th pair was different as 1m + 1sm and 1sm + 1st, respectively. A pair of secondary constrictions were identified in all individuals from different groups investigated in the current study.

Genome size estimation of the sympatric population

In Mt. Seokbyeongsan (pop. 1), we additionally found the tetraploid (2n = 4x = 36) and hexaploid (2n = 6x = 54) in the same place with a few meters apart from each other. Although

it was difficult to clearly distinguish the diploid and tetraploid by morphological differences (Figs. 2a, 6), the hexaploid was clearly recognized by deeply split lobes. And it was confirmed that the chromosome number and genome size are increased according to its ploidy level (Table 4).

DISCUSSION

The *Chrysanthemum zawadskii* complex has been reported in Korea, with ploidy ranging from diploid (2n = 2x = 18) to decaploid (2n = 10x = 90) (Lee, 1967, 1969, 1975; Kim et al. 2004). Diploid can play a role in understanding evolutionary processes, especially in plant groups that underwent polyploidization during speciation. To date, four



Fig. 6. Comparison of the ploidy level and morphological features of tetraploid (A, B) and hexaploid (C, D). *Chrysanthemum zawadskii* distributed together with diploids in the Mt. Seokbyeongsan population. Genome size and ploidy were measured using a CyFlow Ploidy Analyzer (Sysmex Partec, Munster, Germany). Scale bar = 1 cm.

Table 4	. Ploidy	level check	and genome	size estimation	of the	Chrysanthemum	zawadskii	complex in M	t. Seokbyongsan	(Pop.	1)
---------	----------	-------------	------------	-----------------	--------	---------------	-----------	--------------	-----------------	-------	----

No.	Ploidy level (2n)	1C DNA contents (pg)	Genome size (Gbp)	CV (%)
1	2x = 18	3.03 ± 0.01	5.92 ± 0.01	3.41
2	4x = 36	5.71 ± 0.02	11.17 ± 0.04	3.80
3	6x = 54	7.65 ± 0.06	14.97 ± 0.12	3.18

CV, coefficient of variation.

diploid C. zawadskii populations have been reported from two regions in Korea (sites A and B) (Fig. 1) (Kim et al., 2003), in contrast to polyploids that are widely spread over the Korean Peninsula. This was, however, insufficient to explain the evolutionary history of this highly diversified plant group. The current study further identified four new diploid populations of C. zawadskii from natural habitats in Korea (Figs. 2, 4). From the result, it could understand the variation of the Korean C. zawadskii complex in the diploid level together with the already available information of previously reported population. The habitat of some of these new populations was located on mountain peaks, valleys, or ridges of approximately 1,000 m, and they are all included in the Taebaek Mountain Range that runs along the eastern coast of Korea: (1) the diploid population of Mt. Seokbyeongsan was found in a rocky area at the top of the mountain, (2) The population of Mt. Gakhuisan was found in the cracks of the rocks on the cliffs, (3) The population of Mt. Baekisan, a limestone-rich mountain, was sympatric with Chrysanthemum boreale Makino (2n = 18), and (4) The population of Miin Falls was found on the cliffs next to the waterfall in the vicinity of the canyon. These populations are also found in limestonerich areas (Kwon et al., 2002; Song et al., 2016; Kim et al., 2021). The distances between the populations ranged from 24 to 77 km, and the populations of Mt. Gakhuisan and Mt. Baekisan were the closest to each other. These were close to the previously reported site A (Gwangha-ri of Jeongseon-gun and Baekun-ri of Pyeongchang-gun, Gangweon-do, Korea) but distinct from site B (Dalsan and Okgye of Yeoungdeokgun, Gyeongsangbuk-do, Korea) (Fig. 1). In particular, (1) Mt. Seokbyeongsan live together with tetraploids (2n = 4x = 36)and hexaploids (2n = 6x = 54) that is the first report from this study. Each population lived a few meters apart from each other. However, it is difficult to identify the morphological differences between the diploid and tetraploid population (Fig. 6, Table 4). Hexaploid was relatively easily recognized from them by its leaves with finely lobes.

It was revealed that the head flower size of the newly discovered diploid populations ranged from 2.89 to 3.50 cm. This is smaller than previously reported populations of the Korean native *C. zawadskii*, even though their ploidy information was not provided (Kim et al., 2014). The new diploid populations were actually collected before flowering season (except for Miin Falls population), and then transplanted into a greenhouse without ploidy information at first. After stabilizing in the greenhouse condition, it was confirmed the ploidy level using the number of somatic chromosomes. Using these confirmed plant samples, the morphological features were investigated during the flowering

season of the same year. Considering the morphological variations in the genus *Chrysanthemum*, it could suspect the probability of morphological differences between native and greenhouse conditions, but it was not critical limitation to compare the morphological characters among these new diploid populations because they were grown up in the same condition.

Karyotypes of diploid individuals of Chrysanthemum are known to be very diverse even within the same population (Kim et al., 2003). All of the representative karyotype of each population found in this study was included in the previously reported types, however, the number and location of secondary constrictions were differed. It has been generally regarded that structural changes on chromosomes constitute a vital evolutionary mechanism contributing to diversification and speciation, and are especially associated with hybridization and polyploidization in angiosperms (Dowrick, 1952; Rana, 1965; Lee, 1975; Kim et al., 2003; Weiss-Schneeweiss and Schneeweiss, 2012; Kang and Kim, 2020). It is necessary to determine how these diverse karyotypes exist and evolve within the same population and species. It is expected that the discovery of multiple populations of diploid C. zawadskii will provide evidence and suggestion to resolve this existing issue.

From the present study, we found new diploid populations of *Chrysanthemum zawadskii* from the natural habitats of Korea and described the morphological and cytological characteristic. They were all diploids with chromosome number 2n = 18 and diverse karyotypes among the populations. Among them, the sympatric population, where three different ploidy levels are distributed together, was also found at first. It is expected that this finding will contribute to understanding this extensively diverse genera which have experienced the polyploidization and hybridization during their evolution.

ORCID: Mi Jeong MOON https://orcid.org/0009-0004-8444-1870; Sang Hee PARK https://orcid.org/0009-0006-5991-7659; Sung Kyung HAN https://orcid.org/0000-0001-9012-3407; Jung Sung KIM https://orcid.org/0000-0002-7658-6330

ACKNOWLEDGMENTS

This work was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education (2020R111A2053517).

CONFLICTS OF INTEREST

The authors declare that there are no conflicts of interest.

LITERATURE CITED

- Anderson, N. O. 1987. Reclassifications of the genus *Chrysanthemum* L. HortScience 22: 313.
- Dowrick, G. J. 1952. The chromosomes of *Chrysanthemum*, I: The species. Heredity 6: 365–375.
- Fan, E. 1969. Further studies of *Chrysanthemum* breeding for the Southwest. Texas Tech University, Lubbock, TX, 71 pp.
- Heslop-Harrison, J. S. and T. Schwarzacher. 2011. Organisation of the plant genome in chromosomes. The Plant Journal 66: 18– 33.
- Hu, S. Y. 1958. Statistics of Compositae in relation to the flora of China. Journal of the Arnold Arboretum 39: 379–419.
- Kadereit, J. W. and C. Jeffrey. 2007. Flowering Plants, Eudicots: Asterales. The Families and Genera of Vascular Plants, Vol. VIII. Springer, Berlin, 636 pp.
- Kang, J. H. and J. S. Kim. 2020. New diploid populations of *Chrysanthemum indicum* L. (Asteraceae) from Korea. Korean Journal of Plant Taxonomy 50: 17–21.
- Kim, J. H., G. H. Nam, S. B. Lee, S. Shin and J. S. Kim. 2021. A checklist of vascular plants in limestone areas on the Korean Peninsula. Korean Journal of Plant Taxonomy 51: 250–293.
- Kim, J. S., K. Oginuma, K. and Tobe, H. 2008. Analysis of meiotic chromosome behaviour in diploid individuals of *Chrysanthemum zawadskii* and related species (Asteraceae): Evidence for chromosome rearrangements. Cytologia 73: 425–435.
- Kim, J. S., J. H. Pak, B. Seo and H. Tobe. 2003. Karyotypes of metaphase chromosomes in diploid populations of *Dendranthema zawadskii* and related species (Asteraceae) from Korea: Diversity and evolutionary implications. Journal of Plant Research 116: 47–55.
- Kim, J. S., J. H. Pak and H. Tobe. 2004. Chromosome number of *Dendranthema coreana* (Asteraceae). Acta Phytotaxonomica et Geobotanica 55: 63–64.
- Kim, J. S. and H. Tobe. 2009. Variations of leaf thickness in the *Chrysanthemum zawadskii* complex and in two related Korean species: *C. boreale* and *C. indicum* (Asteraceae). Korean Journal of Plant Taxonomy 39: 29–34.
- Kim, S. J., C. H. Lee, J. Kim and K. S. Kim. 2014. Phylogenetic analysis of Korean native *Chrysanthemum* species based on morphological characteristics. Scientia Horticulturae 175: 278–289.
- Kitamura, S. 1978. Dendranthema et Nipponanthemum. Acta Phytotaxonomica et Geobotanica 29: 165–170.
- Kondo, K., C. I. Peng and M. Aoyama. 1999. Chromosome studies in *Chrysanthemum* flora of Taiwan (1) *Dendranthema horaimontana* (Masam.) S. S. Ying and *D. morii* (Hayata) Kitam. Chromosome Science 3: 49–54.
- Kwon, Y. K., S. K. Chough, D. K. Choi and D. J. Lee. 2002. Ori-

gin of limestone conglomerates in the Choson Supergroup (Cambro–Ordovician), mid-east Korea. Sedimentary Geology 146: 265–283.

- Lee, T. B. 2003. Coloured Flora of Korea, Vol I, II. Hayangmunsa, Seoul, 2096 pp. (in Korean)
- Lee, Y. N. 2006. New Flora of Korea. Kyohaksa, Seoul. P. 599.
- Lee, Y. N. 1967. A cytotaxonomic study on *Chrysanthemum zawadskii* Herbich in Korea, 1. Natural hybridization. Korean Journal of Botany 10: 31–35.
- Lee, Y. N. 1969. A cytotaxonomic study on *Chrysanthemum zawadskii* complex in Korea (2) Polyploidy. Korean Journal of Botany 12: 35–48.
- Lee, Y. N. 1975. Taxonomic study on white flowered wild *Chrysanthemum* in Asia. Korean Journal of Botany 14: 63–73.
- Leitch, I. J. and M. D. Bennett. 1997. Polyploidy in angiosperms. Trends in Plant Science 2: 470–476.
- Levan, A., K. Fredga and A. A. Sandberg. 1964. Nomenclature for centromeric position on chromosomes. Hereditas 52: 201–220.
- Liu, P. L., Q. Wan, Y. P. Guo, J. Yang and G. Y. Rao. 2012. Phylogeny of the genus *Chrysanthemum* L.: Evidence from single-copy nuclear gene and chloroplast DNA sequences. PLoS ONE 7: e48970.
- Mahmoudi, S. and G. Mirzaghaderi. 2023. Tools for drawing informative idiograms. *In* Plant Cytogenetics and Cytogenomics: Methods and Protocols. Heitkam, T. and S. Garcia (eds.), Springer, Singapore. Pp. 515–527.
- Makino, T. 1903. Observations on the flora of Japan. Botanical Magazine (Tokyo) 17: 144–208.
- Mayrose, I. and M. A. Lysak. 2021. The evolution of chromosome numbers: mechanistic models and experimental approaches. Genome Biology and Evolution 13: evaa220.
- Oh, I. S., S. O. Yoo and J. H. Bae. 1994. Morphological characteristics of *Chrysanthemum zawadskii* Herb. native to Korea. Journal of the Korean Society for Horticultural Science 35: 78–87.
- Plants of the World Online. 2019. Facilitated by the Royal Botanic Gardens, Kew. Retrieved May 10, 2023, available from: https://powo.science.kew.org/.
- Rana, R. S. 1965. Induced interchange heterozygosity in diploid *Chrysanthemum*. Chromosoma 16: 477–485.
- Song, J. M., H. J. Son, Y. S. Kim, S. C. Kim, D. H. Lee, W. G. Park and S. J. Kwon. 2016. The flora of limestone area, Mt. Seokbyeong. Korean Journal of Plant Resources 29: 241–263.
- Tanaka, R. and N. Shimotomai. 1961. Karyotypes in four diploid species of *Chrysanthemum*. Cytologia 26: 309–319.
- Te Beest, M., J. J. Le Roux, D. M. Richardson, A. K. Brysting, J. Suda, M. Kubešová and P. Pyšek. 2012. The more the better? The role of polyploidy in facilitating plant invasions. Annals of Botany 109: 19–45.

- Trehane, P. 1995. (1172) Proposal to conserve *Chrysanthemum* L. with a conserved type (Compositae). Taxon 44: 439–441.
- Valkonen, J. P. T., K. N. Watanabe and E. Pehu. 1994. Analysis of correlation between nuclear DNA content, chromosome number, and flowering capacity of asymmetric somatic hybrids of diploid *Solanum brevidens* and (di)haploid *S. tuberosum*. The Japanese Journal of Genetics 69: 525–536.

Wang, J. W., J. Yang and M. X. Li. 1993. The morphological vari-

ation and the karyotypical characters of *Dendranthema indicum* and *D. lavandulifolium*. Journal of Systematics and Evolution 31: 140-146.

Weiss-Schneeweiss, H. and G. M. Schneeweiss. 2012. Karyotype diversity and evolutionary trends in angiosperms. *In* Plant Genome Diversity Volume 2: Physical Structure, Behaviour and Evolution of Plant Genomes. Vienna: Springer Vienna. Pp 209–230.